Analysis of the essential oil of *Thymus numidicus* (Poiret) from Algeria

A. Kabouche,¹ Z. Kabouche,¹* and C. Bruneau²

¹ Laboratoire d'Obtention de Substances Thérapeutiques (LOST), Faculté des Sciences, Université Mentouri–Constantine, Campus Chaabet Ersas, 25000 Constantine, Algeria

² Université de Rennes 1, UMR 6509, Campus de Beaulieu, 35042 Rennes Cedex, France

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ABSTRACT: The hydrodistilled essential oil of *Thymus numidicus* (Poiret), grown at Constantine (Algeria), was analysed by GC-MS. The essential oil was largely composed of thymol (68.2%) and carvacrol (16.9%), which are the highest percentages of phenols to be found in an essential oil from a *Thymus* species. Copyright © 2005 John Wiley & Sons, Ltd.

KEY WORDS: Thymus numidicus; Lamiaceae; essential oil; thymol; carvacrol

Introduction

Thymus numidicus (Poiret) (Arabic 'Zaatar') is an endemic species to northern Algeria and Tunisia. The genus Thymus (Family Lamiaceae, Subfamily Nepetoideae, Tribe Mentheae) is represented by more than 300 species; 12 species are distributed in Algeria,¹ nine of them endemic. T. numidicus is characterized by pink flowers and is well known in Algerian folk medicine for its expectorant, antitussive, antiseptic, antispasmodic and anthelmintic properties, as for many other Thymus spp.^{2,3} The antifungal,⁴ antibacterial,⁵ antimicrobial and antioxidant⁶ activities have also been established for Thymus essential oils. Previous investigations on Thymus essential oils revealed an abundance of monoterpene hydrocarbons and phenolic monoterpenes.7-17 The present study deals with the analysis of the essential oil of T. numidicus (Poiret), which is reported for the first time.

Experimental

Plant Material

Dried aerial parts of *T. numidicus* were collected in May 2002 from the mountain Djebel El-Ouahch, Constantine (north-eastern Algeria), at an altitude of ca. 800 m on a clayey soil, by Mr Kamel Kabouche (Constantine) and authenticated by Professor Gérard De Belair (University Badji Mokhtar, Annaba, Algeria). A voucher specimen was deposited at the herbarium of the Laboratory of Therapeutic Substances, Faculty of Sciences, Mentouri-

Constantine University, Constantine, Algeria (LOST ZKAK Tn 05/02).

The hydrodistillation of the dried aerial parts (100 g) of *T. numidicus* for 3 h in a Clevenger-type apparatus, according to the *European Pharmacopoeia*,¹⁸ yielded 2.1% (w/w) of a yellow-orange good-smelling essential oil, which was dried over anhydrous sodium sulphate and, after filtration, stored at +4 °C until tested and analysed.

GC Analysis

GC analyses were performed using a Perkin-Elmer gas chromatograph equipped with two FIDs, a data-handling system and a vapourizing injector port, into which two columns of different polarities were installed: a DB-1 fused silica column ($30 \text{ m} \times 0.25 \text{ mm}$ i.d., film thickness 0.25 µm) and a DB-Wax fused silica column ($30 \text{ m} \times$ 0.25 µm) and a DB-Wax fused silica column ($30 \text{ m} \times$ 0.25 µm) and a DB-Wax fused silica column ($30 \text{ m} \times$ 0.25 µm). Oven temperature was programmed at 45-175 °C at 3 °C/min, then rising at 15 °C/min to 300 °C, then held isothermal (15 min); carrier gas, He at 1 ml/min.

GC chiral analyses were performed using a Perkin-Elmer gas chromatograph equipped with a FID, a datahandling system and a Cyclodex-B fused-silica column ($30 \text{ m} \times 0.25 \text{ mm}$ i.d., film thickness 0.25μ m) and a DB-Wax fused silica column ($30 \text{ m} \times 0.25 \text{ mm}$ i.d., film thickness 0.25μ m). Oven temperature was 75 °C isothermal, injector and detector temperatures, 230 °C and 240 °C, respectively; carrier gas, He at 1 ml/min.

GC–MS Analysis

GC–MS analyses were performed on a Perkin-Elmer apparatus equipped with a DB-1 fused silica column (30 m \times 0.25 mm i.d., film thickness 0.25 µm) and interfaced

^{*} Correspondence to: Z. Kabouche, Laboratoire d'Obtention de Substances Thérapeutiques (LOST), Faculté des Sciences, Université Mentouri– Constantine, Campus Chaabet Ersas, 25000 Constantine, Algeria. E-mail: zkabouche@hotmail.com

with an ion trap detector (ITD; software 4.1). MS operating parameters were as follows: ion trap temperature, 220 °C; split ratio, 1:40; ionization potential, 70 eV; ionization current, 60 μ A; scan range, 40–300 u, scan time, 1 s.

Identification of components was assigned by comparison of the retention indices (RI) relative to C_9-C_{17} *n*-alkanes and MS with corresponding database (NIST library) and with mass spectral literature.^{19–21} Relative percentage amounts of the identified components were calculated from the total ion chromatograms by a computerized integrator.

Results and Discussion

The GC-MS analysis of the yellow-orange essential oil resulted in the identification of 40 components, representing 99.7% of the essential oil. As shown in Table 1, the major components were thymol (68.2%) followed by carvacrol (16.9%) and linalool (11.5%). The predominance of phenolic compounds was in agreement with previous results reported from T. vulgaris essential oil^4 mainly represented with thymol (63.6%), p-cymene (23.5%), γ -terpinene (4.3%) and carvacrol (2.2%) and from T. pectinatus essential oil,6 which was characterized by thymol (49.8%) as the major component, followed by *p*-cymene (14.8%), γ -terpinene (16.1%) and carvacrol (3.7%). In contrast with our results, carvacrol (48.7%)instead of thymol (13.8%) was determined as the main component in the T. serpyllum essential oil.⁵ From our results, the percentages of thymol (68.2%) and carvacrol (16.9%) seem to be the highest amounts of phenols to be found in a *Thymus* species.

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 Table 1. Composition of the essential oil of Thymus numidicus

Compound	Percentage	Retention index
α-Pinene	t	930
Camphene	_	938
1-Octen-3-ol	0.1	961
β -Pinene	t	963
Myrcene	_	975
3-Octanol	_	993
<i>p</i> -Cymene	1.0	1003
Limonene	t	1009
γ-Terpinene	0.3	1035
trans-Linalyl oxide	t	1085
trans-Sabinene hydrate	0.3	1037
cis-Sabinene hydrate	t	1066
Linalool	11.5	1074
Camphor	t	1095
Borneol	0.1	1134
Isoborneol	t	1136
α -Terpineol	0.3	1159
Dihydrocarveol	t	1170
cis-Carveol		1189
Carvone	_	1206
Thymol	68.2	1275
Carvacrol	16.9	1286
Eugenol	t	1327
α-Cubebene		1345
α-Copaene	_	1375
β -Bourbonene	_	1379
α -Humulene	0.3	1447
γ-Muurolene	t	1469
Germacrene D	0.1	1474
β-Silene	t	1485
α -Muurolene	0.1	1494
α -Farnesene	t	1508
δ -Cadinene	_	1522
Caryophyllene oxide	0.3	1561
Spathulenol	_	1577
Unknown	t	1585
Cubenol	_	1600
α -Cadinol	0.1	1626
α -Bisabolol	0.1	1656
Myrcenyl acetate	t	1675
Octadecane	_	1800
Nonadecane	—	1900
Identified components (%)	99.7	
One unknown	t	

t, trace, <0.1%.

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